



Characterization of Mn-induced apoplastic peroxidases in cowpea (*Vigna unguiculata* L.)

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Introduction: Apoplastic H₂O₂-producing peroxidase (POD, NADH-peroxidase) and H₂O₂-consuming POD (guaiacol-POD) are proposed to play key roles in Mn toxicity and Mn tolerance (Fecht-Christoffers et al., 2003). Additionally, apoplastic metabolites significantly affect PODs activities (Fecht Christoffers et al., 2006). In order to characterize the Mn-induced POD isoforms, we separated proteins using blue-native polyacrylamide gel electrophoresis (BN-PAGE). After elution of specific POD isoforms we determined their pH optimum, substrate specificity, and inhibitory effects of different phenols and organic acids.

Results:

1. Several POD isoforms were induced by elevated Mn supply

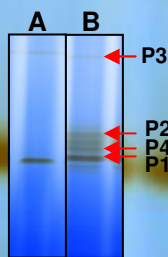


Fig.1. BN-PAGE of apoplastic proteins of the Mn-sensitive cv TVu 91. Plants received either 0.2 μM Mn (A) or were treated with 50 μM Mn for 4 d (B). Marked isoforms were used for further characterization.

3. Only four of nine phenols in varying concentrations affected NADH-peroxidase activity of the examined POD isoforms, but each of them in a different way.

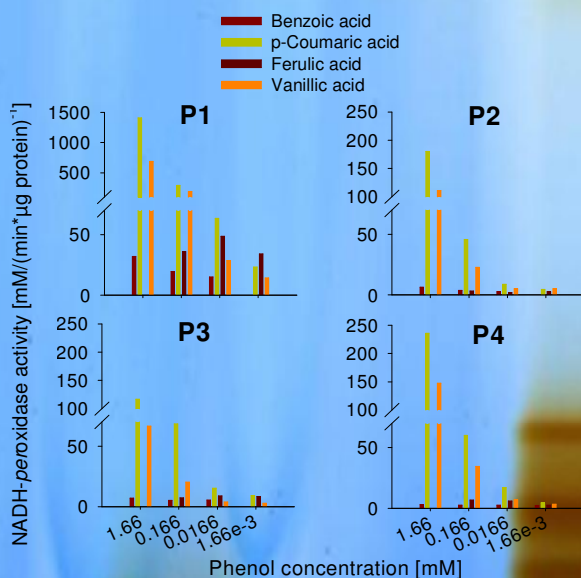


Fig.3. Effect of different phenols and their concentrations on the NADH-peroxidase activity of different POD isoforms.

Conclusions: The fact, that different phenols exerted differential enhancing and inhibitory effects on NADH-peroxidase activities confirm the hypothesis that not only PODs but also apoplastic phenolic metabolites are key factors in developing Mn toxicity and Mn tolerance.

References:

Fecht-Christoffers M. M., Braun, H.-P., Lemaitre-Guillier C., VanDorsseleer A., Horst W. J., 2003, Plant Physiology, 133, 1935-1946
Fecht-Christoffers M.M., Fühns H., Braun H.-P., Horst W.J., 2006, Plant Physiology, 140, 1451-1463
Wehrhahn W and Braun H.-P., 2002, Electrophoresis, 23, 640-646

Materials and Methods: Apoplastic Washing Fluid (AWF) was extracted using a vacuum-infiltration/centrifugation technique (Fecht-Christoffers et al., 2003). Proteins were separated from AWF into protein concentrate and metabolite filtrate using centrifugal concentrators and then separated by BN-PAGE (Fecht-Christoffers et al., 2003). Proteins of interest were eluted from the gels (Werhahn und Braun, 2002) and peroxidase isoform activities were determined spectrophotometrically. Measuring solution consisted of the examined phenol or a combination of phenols, MnCl₂, NADH and sample.

2. Almost each isoform catalysed both, guaiacol-POD and NADH-peroxidase activity. The reaction-dependent pH optima for all isoforms were 6.5 and 5.5, respectively. One isoform (P3) hardly had guaiacol-POD activity.

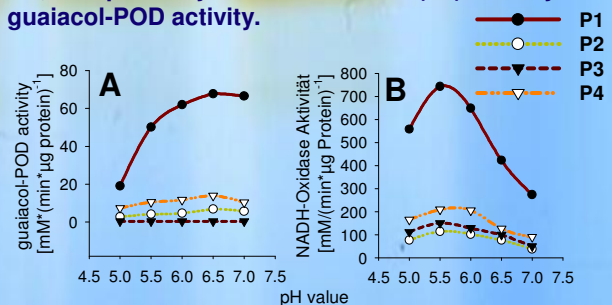


Fig.2. Effect of the pH on the guaiacol-POD (A) and NADH-peroxidase (B) activity of different POD isoforms.

4. Combining p-coumaric acid with different phenols mostly reduced max. NADH-peroxidase activity. Only benzoic acid and vanillic acid enhanced it.

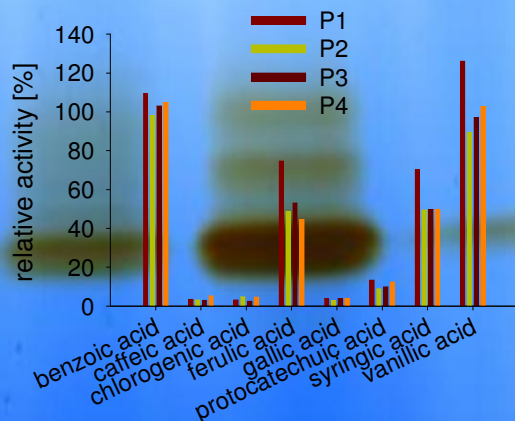


Fig.4. Effect of the different phenols on max. NADH-peroxidase activity of different POD isoforms. Plots are displayed as relative values to the activity only with p-coumaric acid.